

Mantra

How often should I take BF and FL references?	<ul style="list-style-type: none"> Once a week for BF and once a month for FL.
How can I acquire the references?	<ul style="list-style-type: none"> For BF references use a chromogenic slide, focus on the tissue and then move to a clean tissue-free region on the coverslip (large coverslip/small tissue will aid this). For FL references use the blue compensation slide you have received with your Mantra. Acquire the references with the trinocular in shared position and in camera only position.
Which files does Tech Support need from me, and where do I find them?	<p>For instrument related issues:</p> <ul style="list-style-type: none"> This PC \Local Disk (C:)\Users\Public\Akoya\Mantra\Log File <p>For inForm related issues:</p> <ul style="list-style-type: none"> This PC\Local Disk (C:)\Users\Public\Akoya\inForm\Log File
How do I view the FL signal?	<ul style="list-style-type: none"> Check the trinocular position; it should be fully out (camera only) to let the light reach the camera. Check the field stop and aperture stop; they should be fully opened (pushed in). See Figure 1 for more information.
How do I know if I am using BF mode?	<ul style="list-style-type: none"> Check that Köhler illumination is correct. If not please refer to the Mantra user manual.
Contact Technical Support.	<ul style="list-style-type: none"> For technical problem please contact support@akoyabio.com. For application support contact your local FAS. For other information contact CustomerCareEmea@akoyabio.com.
Where can I find more information?	<ul style="list-style-type: none"> Look on our website: www.akoyabio.com. Subscribe to our brand-new blog: www.theinsidescope.blog. Follow us on Twitter, LinkedIn and YouTube.

What do I need to create my own spectral library?

- Singleplex(s):** Library slides establish pure emission spectra for each Opal, as well as DAPI. Each color is stained in singleplex without counterstain. You will also need an “Autofluorescence Only” slide of the same tissue type as your multiplex. This library slide does not take any Opal or counterstaining, but otherwise is treated the same as your multiplex with the same number of antigen retrievals.
- Autoexposure:** Follow the Multispectral Field Band table (Figure 2) to know where you should autoexpose and where not.

FIGURE 1. Trinocular highlighted in blue. Field stop (FP) and aperture stop (AP) highlighted in orange.

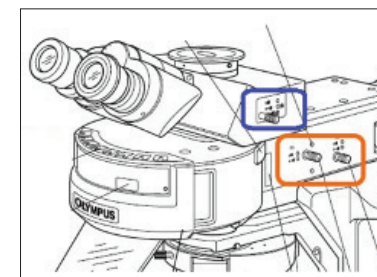


FIGURE 2. Multispectral Field Band table

	Multispectral Field Band						
	DAPI	Opal Polaris 480	FITC	Cy3	Texas Red	Cy5	Opal Polaris 780
DAPI	Colored	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray
Opal Polaris 480	Light Gray	Colored	Colored	Light Gray	Light Gray	Light Gray	Light Gray
Opal 520	Light Gray	Light Gray	Colored	Light Gray	Light Gray	Light Gray	Light Gray
Opal 540	Light Gray	Light Gray	Light Gray	Colored	Light Gray	Light Gray	Light Gray
Opal 570	Light Gray	Light Gray	Light Gray	Light Gray	Colored	Light Gray	Light Gray
Opal 620	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Colored	Light Gray
Opal 650	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Colored
Opal 690	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Colored
Opal Polaris 780	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Light Gray	Colored

KEY LEGEND

COLORED BLOCKS: Signal bands. Autoexpose or manually set an appropriate exposure per sample.

LIGHT GRAY: Non-signal band. Leave the default exposure time.



RESOURCES: Check out our new blog, [The Inside Scope](#).